

The Isomerism of Cobalt(III) Histidinato Chelates

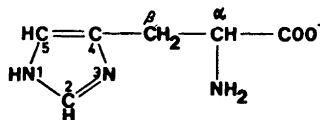
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Circular dichroism and proton magnetic resonance spectra of three isomers of cobalt(III)-L-histidinato complexes have been measured. A new cobalt(III)-DL-histidinato complex has been isolated and characterized.

This study of histidine chelation in monomeric cobalt(III) complexes is part of an investigation of the bonding details in dinuclear, dioxygen-bridged cobalt(III) complexes with histidine as ligand.¹

Fig. 1. Histidinato with the numbering of atoms used. L-Histidine has the (*S*)-configuration in the Cahn-Ingold-Prelog notation.



Histidinato (Fig. 1) has three potential metal binding sites: the amino nitrogen atom, the nitrogen atom number 3 of the imidazole ring, and a carboxylate oxygen atom. In the case of tridentate ligation in an octahedral complex only *facial* attachment is possible, *meridional* chelation being excluded for steric reasons.

Accordingly nine different isomers of the bis(histidinato)cobalt(III) complex with tridentate histidinato may exist as indicated in Fig. 2.

Zompa *et al.*^{2,3} have separated a violet, a red, and an orange isomer of bis(L-histidinato)cobalt(III) and by means of their electronic absorption spectra correlated them to the structures *trans*-carboxylate (3LL), *trans*-imidazole (1LL), and *trans*-amino (2LL), respectively. This assignment was later corroborated by Schmidtke,⁴ who treated Zompas spectra on the basis of the "angular overlap model".

In this work we have confirmed the separation and the electronic spectra of these three $\text{Co}(\text{L-his})_2^+$ isomers* and have measured their CD and PMR spectra.

* $\text{his}^- \equiv \text{histidinato}$

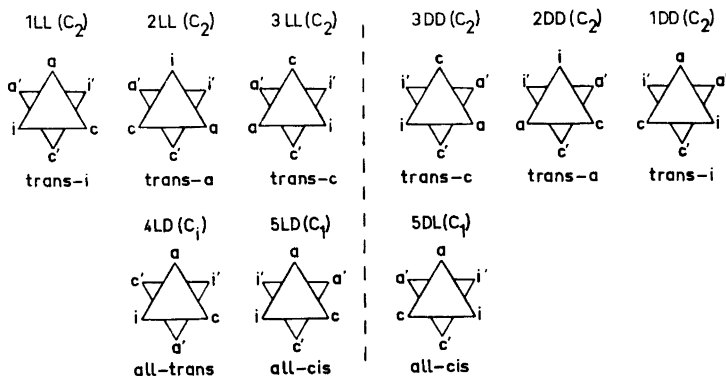


Fig. 2. Isomers of octahedral complexes with two histidinate ligands. L and D denote L- and D-histidinate. a, c, and i indicate the amino, the carboxylate, and the imidazole groups. Dashes are used to distinguish the two ligands. Isomers with the same number are mirror images. The symmetry groups are given in parentheses by the Schoenflies symbols.

We have also studied the complexes formed with racemic histidinate, and have isolated and characterized *rac-all-cis*-[Co(D-his)(L-his)]Br (5DL + 5 LD).

EXPERIMENTAL

Materials. L- and DL-Histidine (*purissimum*, chromatographic purity) were obtained from Fluka. All other chemicals were analytical grade. The cation exchange resin used was 50–100 mesh Dowex 50W-X4 (large effective pore size).

Apparatus. Electronic absorption spectra were measured with a Cary 11 recording spectrophotometer and CD spectra with a Roussel-Jouan Dichrographe II, all measurements at room temperature. Normally PMR spectra were recorded at 60 MHz with a Varian A-60 spectrometer at about 30°C. A 100 MHz Varian HA-100 spectrometer was used for the double-resonance experiments.

Preparations for chromatography. CoCl₂·6H₂O (3.57 g, 15 mmol) and either L- or DL-histidine (5.13 g, 33 mmol) were dissolved in 100 ml water, and 0.5 g activated charcoal was added. Air was bubbled through the solution, while the temperature was maintained at about 75°C. When the colour had changed from brown to red (*ca.* 90 min) the solution was cooled and the charcoal was filtered off.

Separation of isomers. A column of internal diameter 4.1 cm was packed with cation exchanger in the K⁺ form to give a resin bed 15 cm deep. About 50 ml of the solution of isomers, prepared as above, was run on to the column, and eluted with a 0.05 M aqueous KBr solution at a rate of *ca.* 4 ml/min.

When separation had occurred (*ca.* 2 days), the column was allowed to run dry, and the resin containing the individual bands removed, transferred to smaller columns and washed out with 1 M KBr. (4 M KBr was used for the orange component of Co(L-his)₂⁺.)

Brown oxidation products that stayed at the top of the column were always discarded. If required, the resulting KBr containing solutions were concentrated with a rotatory evaporator under reduced pressure; any KBr precipitated during the process was removed by filtration.

The R-values quoted for the isomers are the distance (in cm), travelled by the centre of the band, divided by the volume of eluent (in l) and by the column cross section (in cm²).

Preparation of *rac-all-cis*-[Co(D-his)(L-his)]Br. A solution of Co(III)-DL-histidinate, prepared as for chromatography, was evaporated to half its volume, and an equal volume

of a KBr solution (60 g/100 ml) was added. On standing for some days dark red crystals formed; these were filtered off, washed with cold water and ethanol and dried in the air. Once crystals are at hand, these may advantageously be used for seeding in subsequent preparations. Yield 1.4 g (21 % of total Co). (Found: C 32.21; H 3.61; N 18.86; Br 17.86. Calc. for $[\text{Co}(\text{C}_6\text{H}_8\text{N}_3\text{O}_2)_2]\text{Br}$; C 32.24; H 3.61; N 18.80; Br 17.86.)

PMR Experiments. Solutions of the three fractions of $\text{Co}(\text{L-his})_2^+$ were concentrated to ca. 0.2 M with removal of precipitated KBr. 0.5 ml of these solutions were evaporated to dryness and redissolved in 0.5 ml D_2O . This process was repeated, and the final solution transferred to an NMR tube.

rac-all-cis- $[\text{Co}(\text{D-his})(\text{L-his})]\text{Br}$ was dissolved in 0.5 ml D_2O (35 mg, 0.16 M) evaporated and redissolved in 0.5 ml D_2O .

A pH meter with glass and calomel electrodes was used to get pD values according to the relationship $\text{pD} = \text{pH}_{\text{reading}} + 0.4$.⁵

A trace of *tert*-butylalcohol was added to the tubes, the methyl resonance ($\delta = -1.23$ relative to TMS) being taken as an internal reference; for the double-resonance experiment an external TMS reference was used.

All resonances were observed downfield from TMS giving negative δ -values; the negative sign has been omitted throughout in the following.

Analyses. The elemental analysis was carried out by the Microanalytical Department, Chemistry Laboratory II, University of Copenhagen.

Extinction coefficients of the $\text{Co}(\text{L-his})_2^+$ isomers are based on cobalt analyses. Solutions, containing at least 0.2×10^{-3} mol cobalt, were fumed with conc. H_2SO_4 . Co(II) in the resulting clear solutions was determined by addition of EDTA and back-titration with Zn following the method of Kiss.⁶ The results are estimated to be accurate to within ± 3 %.

RESULTS

Chromatography. The cation-exchange column chromatography of the Co(III)-L-histidinate solution leads to the separation of a violet, a red, and an orange band with R-values 0.08, 0.03, and 0.02, respectively. The mol ratio violet:red:orange was found by absorption measurements to be 3:13:1.

Our attempts to isolate crystals of the three isomers have hitherto been unsuccessful.

The chromatography experiments with Co(III)-DL-histidinate yielded one red band only, having $R = 0.05$. No violet or orange bands were observed. Concentration of the red band eluted with KBr solution resulted in precipitation of dark red crystals with an absorption spectrum identical to that of *rac-all-cis- $[\text{Co}(\text{D-his})(\text{L-his})]\text{Br}$* prepared directly from the reaction mixture.

One chromatographic experiment with Co(III)-DL-histidinate was performed with KCl as the eluting agent in order to get a concentrated PMR sample of the red band without the *all-cis* complex precipitating as the bromide (Spectrum VI, Table 1).

Absorption and circular dichroism. The visible absorption and CD spectra of the KBr containing solutions of the three $\text{Co}(\text{L-his})_2^+$ isomers are given in Fig. 3.

On varying the KBr concentration between 0.16 M and 4 M, the shape of the red isomer absorption spectrum did not change, but the extinction coefficients increased about 4 %.

The absorption and CD spectra of the three isomers were independent of pH in the range 2–6, but as the pH was raised from 6 to 12 a tail from a band in the UV caused an increase in the visible absorption.

Table 1. Data from PMR.

Sample	H_{C_2} peaks		H_{C_5} peaks		$H_{C\alpha}$ peaks		$(H_{C\beta})_2$ peaks		
	δ (ppm)	Pattern ^a	δ (ppm)	Pattern ^a	δ (ppm)	Pattern ^a	δ (ppm)	Pattern ^a	
I	L-Histidine (pD 1.4)	8.75	2(1.35)	7.50	1	4.42	3(6.5)	3.48	2(6.5)
II	Red Co(L-his) ₂ ⁺ (pD 2.5)	7.93	2(1.4)	7.31	2(1.3)	4.00	3(3.7)	3.43	2(3.6)
III	Violet Co(L-his) ₂ ⁺ (pD 5.7)	7.82	2(1.3)	7.41	2(1.1)	4.07	h.o.	3.54	h.o.
IV	Orange Co(L-his) ₂ ⁺ (pD 5.2)	7.93	2(1.5)	7.26	2(1.1)	4.29	h.o.	3.67	h.o.
V	<i>rac-all-cis</i> - Co(D-his) (L-his) ⁺ (pD 6.5)	7.93	2(1.45)	7.38	2(1.4)	4.17	3(3.9)	3.46	2(3.6)
		7.03	2(1.5)	7.15	2(1.4)	3.83	3(3.7)	3.48	2(3.6)
VI	Co(III)-DL-his (mixture) (pD 2.0)	7.95	2	7.45	2	4.26	3	3.51	
		7.06	2	7.20	2	3.86	3		
		7.55		7.31					

^a Number of peaks, and, in parentheses, their separation in Hz.
h.o. ≡ higher order spin patterns.

The dark red crystals of *rac-all-cis*-[Co(D-his)(L-his)]Br dissolved in water showed the absorption spectrum given in Fig. 4 ($\epsilon_{\max} = 87.3$ at 495 nm); its pH dependence was similar to that of the LL-isomers.

Proton magnetic resonance. The 60 MHz spectra in D₂O of L-histidine (I), the three Co(L-his)₂⁺ isomers (II, III, and IV), *rac-all-cis*-Co(D-his)(L-his)⁺ (V), and the chromatographically purified Co(III)-DL-histidinate mixture (VI) are outlined in Table 1. The integration of spectra I–V gave $H_{C_2}:H_{C_5}:H_{C\alpha}:H_{C\beta}$ equal to 1:1:1:2. Addition of a drop of DCl, giving a pD of ~ 0.6 , did not cause significant changes in the spectra reported.

Two independent preparations of *rac-all-cis*-[Co(D-his)(L-his)]Br showed the spectrum in Fig 5; this was analyzed by the double-resonance technique. On irradiating at the resonance frequency of doublet A, B collapsed to a singlet, while C and D remained as doublets. Irradiation of B caused decoupling of A. By irradiation of G, E, and F were decoupled. Irradiation of E and then F revealed that G consists of two nearly superimposed doublets, the higher field doublet arising from coupling with E, the other from coupling with F.

Spectrum VI shows six resonances in the low field region. Four of these correspond closely to those of the *all-cis* isomer, spectrum V. The other two

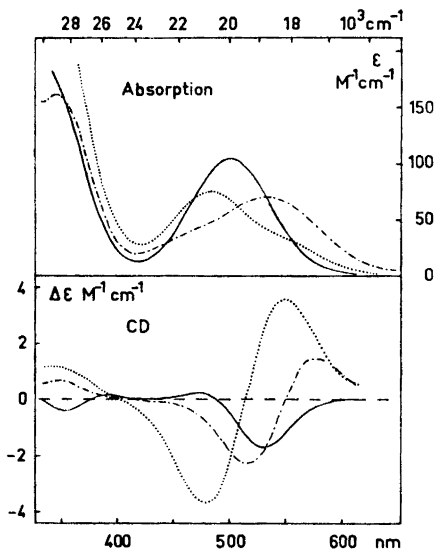


Fig. 3. Solution absorption and CD spectra of the violet (---), red (—), and orange (.....) components of $\text{Co}(\text{L-his})_2^+$ at pH ca. 4. The violet and red solutions were ca. 0.5 M KBr, whereas the orange solution was saturated with KBr.

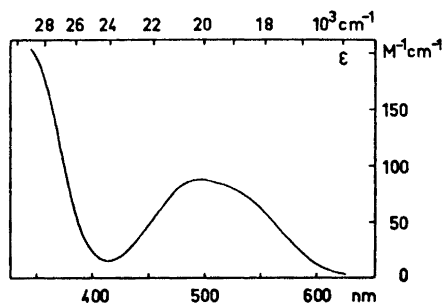


Fig. 4. Solution absorption spectrum of *rac-all-cis*- $[\text{Co}(\text{D-his})(\text{L-his})]\text{Br}$ in H_2O .

(δ values 7.55 and 7.31) were much weaker, their combined intensity being ca. 10 % of the total intensity of all six peaks.

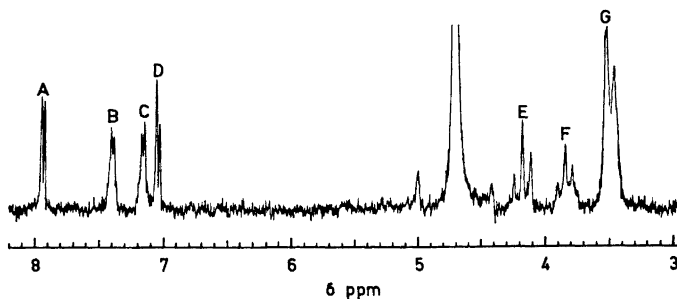


Fig. 5. 60 MHz PMR spectrum of *rac-all-cis*- $[\text{Co}(\text{D-his})(\text{L-his})]\text{Br}$ in D_2O . The ratio of peak areas A:B:C:D:E:F:G is equal to 1:1:1:1:1:1:4.

DISCUSSION

The splitting pattern in the lowest energy spin-allowed *d-d* band was used⁴ in the assignment of structures to the three $\text{Co}(\text{L-his})_2^+$ isomers. The semi-empirical calculations⁴ predict two components in this band of the

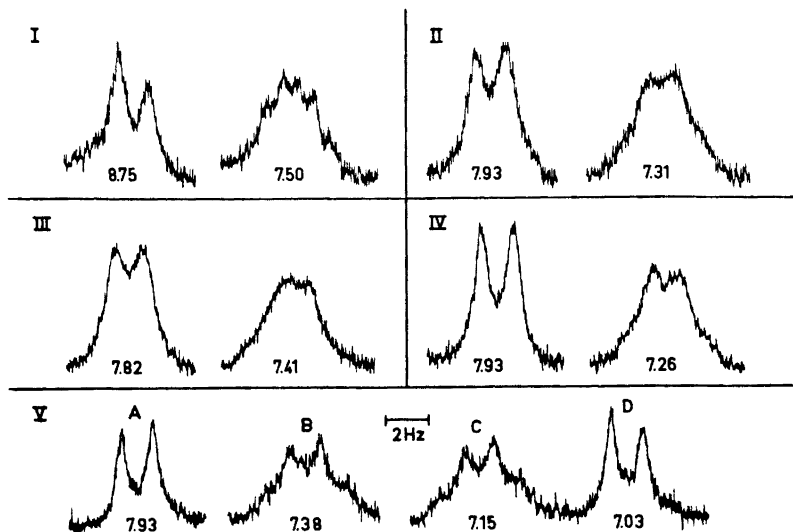


Fig. 6. Expanded scale 60 MHz spectra of H_{C_2} and H_{C_5} protons. δ -value and spectrum number are indicated; compare Table 1 and Fig. 5.

trans-carboxylate and of the *trans*-amino complexes, the calculated patterns being in good agreement with the experimental absorption spectra of the violet and the orange isomers, respectively. As seen in Fig. 3, the CD spectra show a negative and a positive component in the two cases. The *trans*-imidazole structure was expected⁴ to exhibit only a small splitting. This agrees well with the absorption spectrum of the red isomer, which has no discernible splitting; but the CD spectrum reveals that more than one transition occurs, as a negative and a positive component are observed in the first *d-d* band.

The PMR spectra of the diamagnetic complexes are composed of one or more sets of peaks, each of which resembles the spectrum of histidine in acid solution.

The assignments of H_{C_α} and H_{C_β} protons in the spectra of the complexes are straightforward from integration curves and splitting patterns. In histidine itself the resonances of the protons on the imidazole moiety, H_{C_2} and H_{C_5} , have been assigned⁷ as shown in Table 1 on the basis of the pH dependence of their chemical shifts. On an expanded δ -scale it is seen (Fig. 6) that in all spectra the peaks of H_{C_2} and H_{C_5} protons differ in shape, one of them not being a simple doublet. In histidine the broad peak is known to be due to H_{C_5} , and assuming that this is also the case in the complexes we have assigned the H_{C_2} and H_{C_5} peaks as shown in Table 1. The broadening of the H_{C_5} peak possibly arises from coupling with the 2 H_{C_β} protons, in addition to the coupling with H_{C_2} .

The PMR spectra of the $\text{Co}(\text{L-his})_2^+$ complexes are consistent with their C_2 symmetry which implies two equivalent histidinate ligands.

The spectrum of *rac-all-cis*- $[\text{Co}(\text{D-his})(\text{L-his})]\text{Br}$ is composed of two histidine-type spectra. The *all-cis* complexes, structures 5LD and 5DL in Fig. 2, have

C_1 symmetry, and are the only isomers with two non-equivalent ligands. The complex isolated must therefore be a racemic mixture of the two.

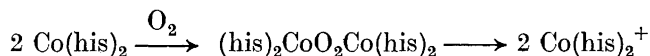
The double resonance experiments make clear that peaks A and B (Fig. 5) originate from the same imidazole ring, and that C and D must therefore arise from the other ligand. We note that the H_{C_2} proton D in the *all-cis* spectrum is situated so far upfield that it appears at higher field than its corresponding H_{C_5} proton, C; this high shielding may be a clue to its precise location in the structure.

The synthesis from racemic histidine could theoretically yield all nine isomers shown in Fig. 2, and so five bands would potentially be separable on an optically inactive resin. In fact we only observe one red band. The absorption spectrum of the eluted band is indistinguishable from that of *rac-all-cis*-Co(D-his)(L-his)⁺; its PMR spectrum, however, does indicate the presence of species other than the *all-cis* isomer, as additional weak peaks are observed (VI, Table 1); and assuming that these are due to the *all-trans* complex (4LD), they account for *ca.* 10 % of the isomers present.

It is clear that DL isomers are predominant in the Co(III)-DL-histidinate preparation. In the labile, octahedral bis(histidinato) complexes of Co(II) and Ni(II) a significant stereoselective effect favouring the mixed DL-metal over the LL- and DD-metal complexes has also been found.⁷⁻¹⁰

It has been suggested⁸ that the imidazole groups in octahedral, bis(histidinato) complexes because of steric hindrance will be forced into *trans* position. The observation of the *all-cis*-, *trans*-carboxylate-, and *trans*-amino-Co(his)₂⁺ isomers shows that this argument does not hold in Co(III) complexes. The *cis*-imidazole arrangement has also been proved in the structure determination of the Co(II) complex [Co(D-his)(L-his)].2H₂O.¹¹

Briefly sketched the route for our preparation of the monomeric Co(III) complexes is



The intermediate μ -peroxo-bis{bis(L-histidinato)cobalt(III)} complex can be obtained in crystalline form.¹² In the light of the foregoing discussion numerous isomers are conceivable of this and similar bridged Co(III) complexes with D- and L-histidine as terminal ligands. Nevertheless it may turn out that stereoselectivity simplifies the picture in reality, as we have found to be the case for the monomeric Co(III) complexes.

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REFERENCES

1. Bagger, S. *Acta Chem. Scand.* **23** (1969) 975.
2. Zompa, L. J., Sokol, C. S. and Brubaker, C. H. *Chem. Commun.* **1967** 701.
3. Zompa, L. J. *Chem. Commun.* **1969** 783.
4. Schmidtke, H.-H. *Chem. Phys. Letters* **4** (1969) 451.
5. Lunry, R., Smith, E. L. and Glantz, R. R. *J. Am. Chem. Soc.* **73** (1951) 4330.

6. Kiss, T. A. *Z. anal. Chem.* **208** (1965) 334.
7. McDonald, C. C. and Phillips, W. D. *J. Am. Chem. Soc.* **85** (1963) 3736.
8. Ritsma, J. H., Van de Grampel, J. C. and Jellinek, F. *Rec. Trav. Chim.* **88** (1969) 411.
9. Morris, P. J. and Martin, R. B. *J. Inorg. Nucl. Chem.* **32** (1970) 2891.
10. Barnes, D. S. and Pettit, L. D. *J. Inorg. Nucl. Chem.* **33** (1971) 2177.
11. Candlin, R. and Harding, M. M. *J. Chem. Soc. A* **1970** 384.
12. Sano, Y. and Tanabe, H. *J. Inorg. Nucl. Chem.* **25** (1963) 11.

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